

In Vitro Anticandidal, Antiviral and Antioxidant Activities of *Cucumis melo L. var. cantalupensis* Naud Extracts

Edziri Hayet^{1,*}, Kaouthar Liouane¹, Fatma Thabti², Fethia Skhiri³, Mahjoub Aouni¹, Maha Mastour¹

¹Laboratoire des maladies transmissibles et des substances biologiquement actives Faculté de Pharmacie-5000-Monastir- Tunisie

²Faculté des sciences Bizerte 7021 Zarzouna, Bizerte

³Laboratory of Genetic Biodiversity and Valorisation of Bioresources, Higher Institute of Biotechnology of Monastir, University of Monastir, Monastir, Tunisia

⁴Laboratoire de Microbiologie C H U Fattouma Bouguiba -5000- Monastir-Tunisia

*Corresponding author: jaziri_hayet@yahoo.fr

Abstract Anticandidal, antiviral and free radical scavenging effects of aerial part and flesh extracts of *Cucumis melo L. var. cantalupensis* were investigated. Total phenolic content of extracts were determined using Folin–Ciocalteu method. The anticandidal activity was evaluated using microwell dilution method against four fungi. The antiviral activity was determined against human cytomegalovirus (HCMV) strain AD-169 (ATCC Ref. VR 538) using a cytopathic effect (CPE) reduction assay. Antiradical scavenging capacities of *Cucumis melo* extracts were tested using free radical forms of ABTS. Among tested extracts, aerial part extracts showed the best anticandidal activity with Minimal Inhibitory Concentration (MIC) ranged from 0.256 to 2.5 mg/ml and Minimal Fungicidal Concentration (MFC) ranged from 2.5 to 5 mg/ml. In addition, such extracts exhibited the highest antiviral and antiradical activities. The results provided an evidence that the studied fruit might, indeed, be potential sources of natural antioxidant and antimicrobial agents.

Keywords: *Cucumis melo L.*, Anticandidal, Antiviral, ABTS, Activity

Cite This Article: Edziri Hayet, Kaouthar Liouane, Fatma Thabti, Fethia Skhiri, Mahjoub Aouni, and Maha Mastour, “In Vitro Anticandidal, Antiviral and Antioxidant Activities of *Cucumis melo L. var. cantalupensis* Naud Extracts.” *Journal of Food and Nutrition Research*, vol. 4, no. 9 (2016): 596-599. doi: 10.12691/jfnr-4-9-6.

1. Introduction

Plants play a significant role in maintaining human health and improving the quality of human life. They serve humans well as valuable components of food, such as seasonings and beverages as well as in cosmetics, dyes, and medicines. Many plant extracts prepared from plants have shown to exert biological activity in vitro and in vivo, which justified research on traditional medicinal plants focused on the characterization of their antimicrobial activity [1]. Large numbers of plants have been screened as a viable source of natural antioxidants including tocopherols, vitamin C, carotenoids and phenolic compounds which are responsible for maintenance of health, to help the human body to reduce oxidative damage and to protect from coronary heart diseases and cancer [2,3]. Phytochemicals in fruits and vegetables can neutralize oxidative agents. Beneficial effects of phytochemicals are believed to be achieved through several mechanisms, such as stimulation of the immune system, modulation of gene expression and hormone metabolism, chelation of transition metals and providing antibacterial and antiviral supports. The health benefits of vegetables in preventing cancer and cardiovascular diseases are mostly attributed to the quality and quantity

of antioxidative components. The Cucurbitaceae family includes several species of cultivated plants of great economic importance, including watermelon (*Citrullus lanatus L.*), squash (*Cucurbita maxima L.*), cucumber (*Cucumis sativus L.*) and cantaloupe (*Cucumis melo L.*) [4]. Cantaloupe is one of the most consumed fruit crops worldwide due to its pleasant flavor and nutritional value. Cantaloupes are a diverse group of fresh, dessert fruits that includes the orange flesh cantaloupes, green flesh honeydew, and mixed melons. Other studies showed that cantaloupe pulp extracts possesses antioxidant and anti-inflammatory properties [5,6].

The object of this study was to determine the anticandidal, antiviral and free radical scavenging activities of the aerial part (leaf and stem) and fruit extracts of *Cucumis melo* growing in Tunisia (Kerker).

2. Materials and Methods

2.1. Plant Material

The herb was purchased in June from a local market in Kerker (sahel Tunisia) and the plant aerial parts and fruit were authenticated and a voucher specimen was deposited in our laboratory of Faculty of Pharmacy.

2.2. Preparation of Extracts

2.2.1. Ethanol extract

Each sample (50 g) of flesh and aerial parts (stem and leaf) was incubated with 200 ml of ethanol (80%) for 3 days under magnetic stirrer. Solvent was evaporated under vacuum at 70°C to get crude extracts and it was stored at -80°C until use.

2.2.2. Aqueous Extract

Each sample (50 g) of aerial parts and flesh of *Cucumis melo* L was extracted with water at 80°C, for 30 min under continuous shaking. The extract was filtered using a Whatman no 1 filter paper. The water extracts were stored at -80°C prior to experimentation.

2.3. Total Phenolic Contents

The total phenolic content in each extract was determined using Folin – Ciocalteus reagent according to the method of Singleton and Rosi [7]. Forty microliters of extract (1 mg/ml) were mixed with 200 µl Folin – Ciocalteus reagent (Sigma–Aldrich, Germany) and 1160 µl of distilled water, followed by 600 µl 20% sodium carbonate (Na_2CO_3) 3 min later. The mixture was shaken for 2 h at room temperature and absorbance was measured at 765 nm. All tests were performed in triplicate. Catechin (Sigma – Aldrich, Germany) was used as a standard. The concentration of total phenolic compounds (TPC) was determined as mg Catechin Equivalent (CE) per gram extract.

2.4. Determination of Anticandidal Activity

2.4.1. Fungi

The antifungal effect of the extracts was also tested against a range of pathogenic reference yeasts: *Candida albicans* ATCC 90028, *Candida glabrata* ATCC 90030, *Candida kreusei* ATCC 6258 and *Candida parapsilosis* ATCC 22019.

2.4.2. Determination of Anticandidal Activity

The antimicrobial activity of the extracts was evaluated through the determination of the minimal inhibitory concentration (MIC) by the micro well dilution method [8]. All extract stock solutions were prepared by dissolution in 10% dimethyl sulfoxide (DMSO). The tested plant extract concentrations ranged from 1 to 10 mg/ml. The MIC of each extract was defined as the lowest concentration which inhibited candidal growth, after incubation at 37°C between 18 and 24h. The minimal fungicidal concentration (MFC) was determined by subculture on blood agar at 37°C between 18 and 24 h.

2.5. Antiviral Activity

2.5.1. Cell Toxicity Assay

The evaluation is based on the reduction of MTT (3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide). The MTT colorimetric assay was performed in 96-well plates [9]. Human diploid embryonic lung fibroblasts (MRC -5) cells were seeded in 96-well plates at a

concentration of 10^5 cells per well and incubated for 24 h at 37°C in a 5% CO_2 enriched atmosphere. After treatment with various concentrations of each extract, cells were incubated for an additional 48 h at 37°C. After that, medium was removed, the cells in each well were incubated with 200 µL of MTT solution (5 mg mL^{-1}) for 2 h at 37°C. MTT solution was then discarded and 200 µL insoluble formazan crystal was added. The optical density (OD) was measured at 540 nm. Data were obtained from triplicate wells. The cytotoxic concentration of the compound was expressed as IC_{50} , the concentration of the tested material required to kill the cells by 50%.

2.5.2. Test Viruses

Human cytomegalovirus (HCMV) strain AD -169 (ATCC Ref. VR 538) was grown on MRC-5 cells in MEM medium until complete cytopathic effect (CPE). The titer viral was used at a final concentration of 100TCID_{50} (50% Tissue Culture-Infective Dose) which were determined by the method of Reed and Muench [10].

2.5.3. Antiviral Activity Assay

A CPE reduction assay for screening the antiviral activities of the plant extracts was employed. In brief, 100TCID_{50} (50% tissue culture-infective dose) virus suspension and serial two-fold dilutions of crude extracts were added simultaneously to confluent cell monolayers in a 96-well plate. The dilution medium without samples and with virus suspension were respectively added, to the cell cultures to serve as cell control and virus control. The plates were incubated at 37°C in a humidified CO_2 atmosphere for 3–5 days. The concentration that reduced 50% of CPE compared to the virus control was estimated from the plots of data and was defined as the 50% inhibitory concentration (IC_{50}). The selective index (SI) was calculated from the ratio $\text{CC}_{50}/\text{IC}_{50}$ [11].

2.6. Radical Scavenging Activity

2.6.1. Radical Cation ABTS+• Scavenging Activity

The standard method described by Dorman and Hiltunen [12] was adopted with minor modifications. This assay assesses the total radical scavenging capacity based on the ability of a compound or an extract to scavenge the stable ABTS radical $\text{ABTS}^{+\bullet}$. The blue-green ABTS radical form was produced through the reaction between ABTS and potassium persulfate in water. A concentrated $\text{ABTS}^{+\bullet}$ stock solution was diluted with phosphate buffered saline (PBS) at pH 7.4 to a final absorbance of 0.7 ± 0.02 with a wavelength of 734 nm and at a temperature of 25°C. Solutions with different diluted concentrations of our samples (extracts and natural products) were prepared in ethanol. Ten microliters of an antioxidant-containing solution were added to 990 µl of $\text{ABTS}^{+\bullet}$ solution and the absorbance was measured at 734 nm. Sample Absorbance was compared to a blank where 10 µl of the solvent were added to 990 µl of the $\text{ABTS}^{+\bullet}$ solution. Absorbance was measured at 20 minutes after addition of the antioxidant. All measurements were performed in triplicate. Results were expressed as percentage inhibition.

3. Result and Discussions

3.1. Total Phenolic Content

Phenolic compounds in plants constitute a major class of secondary plant metabolites with bioactive potential attributed to antioxidant and antibacterial activities. The Total phenolic content (TPC) was expressed in mg catechin equivalent per gram of extract (mg CE/g of extract). The results of the total phenolic content of *Cucumis melo* extracts were given in Table 1. The total phenolic content varied from 10.15 to 75.34 mg CE/g of extracts. The results indicate that the ethanolic extract of aerial parts of *C. melo* had the highest total phenolic content (75.34 mg CE/g of extracts) whereas the lowest content was measured in the aqueous flesh extract (10.15 mg CE/g of extracts). The result presented in Table 1 illustrates the efficiency of ethanol for the extraction of total phenolic compounds. Phenols are very important plant constituents because of their radical scavenging ability due to their hydroxyl groups [13]. This finding is in agreement with some previous studies which reported that the total phenolic content of leaf extract is higher than in other parts of the plant for *Beta vulgaris*, *Petroselinum crispum* and *Coriandrum sativum* [14,15]. This suggests that leaf might be the part that is rich in phenolic compounds in many plants.

Table 1. Total phenolic content of *Cucumis melo* L. extracts

Cucumis melo part	Extract types	Total phenolic content
Aerial	Aqueous	25.21±1.2
	Ethanol	75.34±2.2
flesh	Aqueous	10.15±1.5
	Ethanol	15.58±0.15

Values are given as means ± SD; total phenolic content (mg CE/g) is given in mg catechin equivalent/g extract.

3.2. Anticandidal Activity

All the extracts tested from Tunisian *Cucumis melo* showed anticandidal activity against all tested fungi. MIC ranged from 0.256 to 2.5 mg/ml and MFC ranged from 2.5 to 5 mg/ml (See Table 2). The strongest inhibitions were obtained with ethanolic extract of aerial parts with MIC of 0.256 mg/ml and MFC of 2.5 mg/ml. The aqueous extract of aerial parts of *C. melo* showed also anticandidal activity with MIC of 0.512 mg/ml. Moderate anticandidal activity was also observed with flesh extracts. The anticandidal activity might also be attributed to the high quantity of polyphenols, which are known to possess efficient antimicrobial activity [16]. In other works phenolic compounds have been reported to be responsible for antimicrobial properties [17].

Table 2. Anticandidal activity of *Cucumis melo* L. extracts using microwell dilution method

Extracts	<i>C. glabrata</i> ATCC 90028	<i>C. albicans</i> ATCC 90030		<i>C. krussei</i> ATCC 6258		<i>C. parapsilosis</i> ATCC 22019			
		MIC ^a	MFC ^a	MIC ^a	MFC ^a	MIC ^a	MFC ^a	MIC ^a	MFC ^a
Aerial parts	Et	0.256	2.5	0.256	2.5	0.256	2.5	0.256	2.5
	Aq	0.512	2.5	0.512	2.5	0.512	2.5	0.512	2.5
Flesh	Et	2	5	2.5	5	2.5	5	2.5	5
	Aq	2	5	2.5	5	2.5	5	2.5	5

^a Results are means of six different experiments (n=6).

Et: ethanol extract; Aq: aqueous extract; MIC: minimal inhibitory concentration; MFC: minimal fungicidal concentration; values given as mg/ml.

Table 3. Antiviral activity of *Cucumis melo* L. extracts

Extracts	Anti-HCMV			
	IC ₅₀ (µg mL ⁻¹) ^a	CC ₅₀ (µg mL ⁻¹) ^b	SI ^c	
Aerial parts	Aqueous	150	>300	>2
	Ethanol	100	>300	>3
Flesh	Aqueous	250	>300	>1,2
	Ethanol	250	>300	>1,2
Ganciclovire ^d		0,8	>200	>250

^aIC₅₀ is the concentration of the sample required to inhibit 50% virus-induced CPE.

^bCC₅₀ is the concentration of the 50% cytotoxic effect.

^cSI (selective index) is the ratio CC₅₀/IC₅₀.

^dGanciclovire which are clinically used anti-HCMV drugs was used as positive control in the antiviral activity.

3.3. Antiviral Activity

The antiviral activity was estimated on the basis of the cytopathic effect (CPE) of the virus-infected confluent monolayer of MRC 5 cells. The mean IC₅₀, CC₅₀ and SI values are given in Table 3. All extracts were not toxic against MRC5 cells (CC₅₀>300 µg mL⁻¹). The most active extracts were ethanol and aqueous extracts of aerial

parts of *C. melo*, which inhibited HCMV virus replication at 100 and 150 µg/ml without showing cytotoxic effects and with a selective index higher than 3 for the ethanolic extract. Good antiviral activity was also found with flesh extracts. The observed antiviral activity may be due to the higher amount of phenolic compounds particularly flavonoids and tannins known to possess good antiviral activities [18]. It was reported that extracts from rosemary and provenci al herbs showed potential antioxidant and anti - HIV activities [19].

3.4. Radical Scavenging Activity

Compared with Trolox, the maximal inhibition percentage values calculated after 20 minutes of reaction showed different antiradical activities for a l l four tested extracts. The ethanol aerial parts extract shows the best ABTS inhibition with IC₅₀ of 8.16 mg/ml. The aqueous aerial parts extract also showed good antiradical activity with IC₅₀ of 10.12 mg/ml. Aqueous flesh extract showed a moderate inhibition with a IC₅₀ value of 20.21 mg/ml. The observed antioxidant activity may be explained by the total phenolic content in the active extracts. A high correlation between total phenolic content and antioxidant activity was reported in different studies [20,21].

Table 4. ABTS inhibition percentage of *Cucumis melo L.* extracts

	Extracts	IC50(mg/ml)
Aerial parts	Aqueous	10.12±2.5
	Ethanol	8.16±1.32
Flesh	Aqueous	20.21±0.56
	Ethanol	17.56±1.2
Trolox		0.122±0.02

IC50 (mg/ml) concentration scavenging 50% of ABTS free radicals.

4. Conclusion

The present study shows that ethanolic and aqueous extracts of aerial parts of *Cucumis melo L.* had the highest total phenolic content. It also showed the best anticandidal activity. Furthermore, the ethanolic aerial parts extract showed strong radical scavenging activity against ABTS radical and an important antiviral activity against HCMV. Thus, these extracts can be considered as potential new sources of natural antioxidants for food and nutraceutical products. At present it is not yet established what components are responsible for the observed activities, further work should therefore be performed on the isolation and identification of the active compounds.

References

- [1] Martinez, G., Delgado, R., Pérez, G., Selles, A., Leo, S. Evaluation of the in vitro antioxidant activity of *Mangifera indica L.* extract (Vimang). *Phytotherapie Research*, 14(3). 424-427. 2000.
- [2] Yanga, J.H., Linb, H.C., Maub, J.L. Antioxidant properties of several commercial mushrooms. *Food Chemistry*, 77(2). 229-235. 2004.
- [3] Halliwell, B., Gutteridge, J.M.C. *Free Radicals in Biology and Medicine*. Oxford, 2007, 225p.
- [4] Ritschel, P.S., Lins, T.C., Tristan, L., Buso, G.S.C., Buso, J.A., Ferreira, M.E. Development of microsatellite markers from an enriched genomic library for genetic analysis of melon (*Cucumis melo L.*). *BMC Plant Biology*, 4 (2). 9. 2004.
- [5] Ismail, H.I., Chan, K.W., Mariod, A.A., Ismail, M. Phenolic content and antioxidant activity of cantaloupe (*Cucumis melo*) methanolic extracts. *Food Chemistry*, 118(2). 120-127. 2010.
- [6] Vouldoukis, I., Conti, C., Krauss, P., Kamaté, C., Blazquez, S., Tefit, M., Mazier, D., Calenda, A., Dugas, Q. B. Supplementation with gliadin-combined plant superoxide dismutase extract promotes antioxidant defences and protects against oxidative stress. *Phytotherapie Research*, 18(12). 957-962. 2004.
- [7] Singleton, V.L., Rosi, J.A. Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *American Journal of Enologie and Viticulture* 16(2). 144-58. 1965.
- [8] Smania J., Smania, A., Delle Monache, F., Pizzolatti, M., Delle Monache, G. Derivatization does not influence antimicrobial and antifungal activities of applanoxidic acids and sterols from *Ganoderma spp.* *Zeitschrift für Naturforschung B*, 61(6). 31-34. 2006.
- [9] Polydoro, M., De Souza, K.C.B., Andrades, M.E., Da Silva, E.G., Bonatto, F., Heydrich, J., Dal-Pizzol, F., Schapoval, E.E.S., Bassani, V.L., Moreira, J.C.F. Antioxidant, a pro-oxidant and cytotoxic effects of *Achrocline saturoioides* extracts Brazil. *Life Science*, 74(23). 2815-2826. 2004.
- [10] Reed L.J., Muench H. A simple method of estimating fifty percent endpoints. *American Journal of tropical medicine and Hygiene*, 27(20). 493-497. 1938.
- [11] Kujungiev, A., Tsvetkova, I., Serkedjieva, Y., Bankova, V., Christov, R., Popov, S. Antibacterial, antifungal and antiviral activity of propolis of different geographic origin. *Journal of Ethnopharmacology*, 64(3). 235-240. 1999.
- [12] Dorman, H. J. D., Hiltunen R. Fe(III) reductive and free radical-scavenging properties of summer savory (*Satureja hortensis L.*) extract and subfractions. *Food Chemistry*, 88(3).193-199. 2004.
- [13] Peter, Y.Y., Wong, D.D. Studies on the dual antioxidant and antibacterial properties of parsley (*Petroselinum crispum*) and cilantro (*Coriandrum sativum*) extracts. *Food Chemistry*, 97(4). 505-515. 2006.
- [14] Pyo, Y.H., Lee, T.C., Logendra, L., Rosen, R.T. Antioxidant activity and phenolic compounds of Swiss chard (*Beta vulgaris* subspecies *cycla*) extracts. *Food Chemistry*, 85(4).19-26. 2004.
- [15] Wong, P.Y.Y., Kitts, D.D. Studies on the dual antioxidant and antibacterial properties of parsley (*Petroselinum crispum*) and cilantro (*Coriandrum sativum*) extracts. *Food Chemistry*, 97(4). 505-515. 2006.
- [16] Edziri, H. Smach, M.A., Ammar, S., Mahjoub, M.A., Mighri Z., Aouni, M., Mastouri, M. Antioxidant, antibacterial, and antiviral effects of *Lactuca sativa* extracts. *Industrial Crops and product* 34(3). 1182-1185. 2011.
- [17] Chun, O. K., Kim, D. O., Smith, N., Schroeder, D., Han, J. T. , Lee, C. Y. Daily consumption of phenolics and total antioxidant capacity from fruits and vegetables in the American diet. *Journal of Science Food and Agriculture*, 85(3).1715-1724. 2005.
- [18] Namba, T., Shiraki, K., Kurokawa, M. Development of antiviral agents from traditional medicines. In: Ageta, H. Aimi, N. Ebizuka, Y. Fujita, T. Honda, G. (Eds.), *Towards Natural Medicine Research in the 21st Century*. Elsevier Science, Amsterdam, 1998.
- [19] Aruoma, O.I., Spencer, J.P., Rossi, R., Aeschbach, R., Khan, A., Mahmood, N., Munoz, A., Murcia, A., Butler, J., Halliwell, B. An evaluation of the antioxidant and antiviral action of extracts of rosemary and provencal herbs. *Food Chemical Toxicology*, 34(2). 449-456. 1996.
- [20] Edziri, H., Smach, M.A., Ammar, S., Mahjoub, M.A., Mighri, Z., Aouni, M., Mastouri, M. Antioxidant, antibacterial, and antiviral effects of *Lactuca sativa* extracts. *Industrial Crops product* 34(2). 1182-1185. 2011.
- [21] Mahjoub, M., Ammar, S. Edziri, H. Mighri, N. Bouraoui, A., Mighri, Z. Anti-inflammatory and antioxidant activities of some extracts and pure natural products isolated from *Rhus tripartitum* (Ucra). *Medicinal Chemistry Research* 19(2).271-282. 2004.