

Detection and Molecular Characterization of Respiratory Syncytial Virus (RSV) in Children with Respiratory Signs in Khartoum State, Sudan 2011-2012

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Abstract Background: The present study was to investigate the incidence of the respiratory syncytial virus infection in children and to characterize the RSV circulating in Khartoum state during 2011-12 winter seasons. **Methodology:** Throat swab specimens collected from 224 children less than 5 years old, with respiratory tract infections admitted at Khartoum Hospitals in winter season (2011- 2012), were screened for RSV using direct immunofluorescence assay (DFA) and reverse transcription- polymerase chain reaction (RT-PCR). Isolation in cell culture followed by nucleotide sequencing and bioinformatics analysis based on the G gene, were done for the RT-PCR positive RSV samples. **Results:** Out of 224 patients, RSV infections were detected in 136 (60.7%) patients, by using DFA technique, and 44 (19.6%) patients using RT-PCR. 22 strains of RSV were isolated in Hep-2 cell line. The clinical symptoms including Bronchiolitis, Pneumonia, Asthma and Allergy showed significantly different rates ($p < 0.05$) in having RSV infection, (P -value = 0.017, 0.002, 0.0001, 0.0001) respectively. Bioinformatics analysis of nucleotide sequences of 7 cell culture isolated RSV strains revealed that all analyzed RSV belonged to the RSV-A genotype. Phylogenetic tree of RSV-A sequences showed that, all Sudanese strains were grouped with strains from Belgium and Saudi Arabia. **Conclusions:** This is the first report on molecular characterization that describes the circulation of RSV genotype in Sudan. DFA and RT-PCR offers rapid methods for detection of RSV in hospitalized children with Respiratory tract infection (RTI).

Keywords: respiratory tract infection, respiratory syncytial viruses, reverse transcription polymerase chain reaction, direct immunofluorescence assay, khartoum

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1. Introduction

Respiratory syncytial virus (RSV) is the most common cause of bronchiolitis and pneumonia among infants and children, particularly during the first year of life [1]. RSV is a negative-stranded non-segmented RNA virus in the genus Pneumovirus, subfamily Pneumovirinae of the family Paramyxoviridae [2]. The majority of children hospitalized for RSV infection are under 6 months of age [3]. RSV infections are responsible for 27-96% of hospitalized cases in developing countries [4], and about 100,000 infants are hospitalized annually in the United States alone [5,6].

RSV infections are also responsible for mortality, with almost nine times the mortality rate of influenza [7]. Developing countries had the highest mortality rates, with

an estimated 66,000 to 199,000 deaths occurring in children younger than five years of age in 2005 [7].

RSV infections in temperate climates usually commence during the winter season, from October to December, as epidemics that may last for up to 5 months [8,9]. Although RSV has been recognized as an important pathogen, unfortunately no infant vaccine or antiviral treatment is presently available against RSV infections [10,11]. RSV subtypes A and B, are present in most outbreaks. Subtype A usually causes more severe disease [12,13], and is usually the predominantly circulating strain compared with subtype B strain [14,15].

Laboratory methods currently available for the detection of RSV include, virus isolation in cell culture, detection of viral antigens by direct or indirect immunofluorescent (IF) staining (DFA/IFA) or by enzyme-linked immunosorbent assays (ELISA) and the detection of viral nucleic acids by amplification assays

[16], mainly reverse transcription polymerase chain reaction (RT-PCR) which represent a rapid and sensitive method for detection of RSV compared to the other techniques [17].

In Sudan work on RSV is very sparse, little information is available about the Incidence of RSV infection in infant, however pneumonia was the first diseases leading to death in 2004, 2005 and 2006 in infants in Sudan [18]. We previously showed an overall sero-prevalence rate of RSV infection in 24 out of 92 (26%) infants with RTI. The sero-prevalence was higher in the 1-2 age groups (unpublished data).

The present study aimed to investigate the incidence of respiratory syncytial virus (RSV) infection in Khartoum state from Gaffer Ebinauff and omdorman children hospitals, in infants and children under 5 years of age with respiratory tract infections (RTI) during 2011-12 winter seasons. The epidemiological data and molecular characterization of detected RSV genotype were analyzed.

2. Materials and Methods

2.1. Study Area

The study was conducted in Khartoum State of Central Sudan, Patients involving children under 5 years of age with respiratory tract infection (RTI) seen at the emergency department in Khartoum Hospitals (Gaffer Ebinauff and omdorman children hospitals), were recruited between the 1st of January to the end of March in 2011 and 2012.

2.2. Data Collection

Demographic data of the patients were collected using a structured questionnaire, which included the following criteria: Age, gender, and clinical symptoms (pneumonia, bronchiolitis, history of asthma and allergy). The specimens were collected within 1-7 days of disease onset.

2.3. Collection of Specimens

A total of 224 throat swab samples (112 samples in each year) were collected from patients by using sterile nylon swabs (Regular Flocked swab, Cat. No. 520CS01, Copan Diagnostics Inc., Murrieta, Calif, USA) in 3 ml of transport media (UTM-RT, Cat. No. 92562, Copan Diagnostics Inc., Murrieta, Calif, USA). Samples collected were transported on ice at the same day of collection to the Department of Virology, Central Laboratory (Ministry of Science and Technology) and stored at -80°C until processed.

2.4. Direct Immunofluorescence Assay (DFA)

Slides for DFA were prepared according to the manufacturer's instructions (ARGENE Respiratory Syncytial Virus DFA kit, Varilhes, France), three times wash with PBS was done for the direct specimens, in which 5 ml of PBS was added to 0.5 ml of the secretions, pipetted gently and centrifuged at 180 g (1.200 rpm) for 10 min at 4°C . Then 30 μL of the suspension sediment was obtained on a slide for immunofluorescence purposes,

finally dried then fixed using chilled acetone for 10 min and stored at -20°C for further work.

DFA was carried out using specific fluoresce in conjugated monoclonal antibody (ARGENE Respiratory Syncytial Virus DFA kit, Varilhes, France) in accordance with the manufacturer's instructions, for the detection and identification of RSV antigen in direct respiratory specimen.

2.5. RNA extraction and cDNA synthesis

RNA was extracted using the Ribo-prep nucleic acid extraction kit (Ecolis.r.o., Bratislava, Slovak Republic), in accordance with the manufacturers protocol. The RNA was extracted from 100 μL of specimen and 10 μL of internal control was added to each sample. cDNA synthesis was performed using Reverta-L reagents kit, according to the protocol of the manufacturer (Ecolis.r.o., Bratislava, Slovak Republic) under the following conditions: The tested tubes were placed in the thermocycler and incubated at 37°C for 30 min. The cDNA samples were diluted by adding 20 μl from DNA-buffer.

2.6. RT-PCR for RSV

The reverse transcription- polymerase chain reaction (RT-PCR) assay targeting G gene was performed using a commercial kit following the manufactures instructions (hRSV-Eph-PCR kit, Ecoli, Slovak Republic). The reaction was conducted in a thermocycler with block temperature adjustment, using the following protocol; 95°C for 5 min for initial denaturation, 42 cycles of 95°C for 45 s, 56°C for 45 s, and 72°C for 45 s, final extension at 72°C for 1 min. All the amplified products were subjected to 1.5% agarose gel electrophoresis. Positive specimens for RSV resulted in PCR fragments of 298 bp and the internal control resulted in PCR fragment of 550 pb.

2.7. Virus Isolation

Viral culture was done to the positive RSV samples by RT- PCR. Human epithelial type 2 (HEp-2) was obtained from (Vircell, Granada, Spain) and was grown at 37°C in 5% CO_2 in modified Eagle's medium supplemented with 10% fetal bovine serum (FBS). 1 ml of each sample was inoculated in HEp-2 monolayer (60%–70% confluent) cultures and was maintained in modified Eagle's medium supplemented with 2% fetal bovine serum and 1% penicillin-streptomycin. Following adsorption, fresh medium was added, and inoculated cells were observed daily for cytopatic effect (syncytia) for up to 7 days. Virus was harvested when the monolayer demonstrated approximately 75% CPE (usually on days 3 or 4 post-inoculation). The cells were subjected to three successive freeze-thaw cycles followed by re-suspension in fresh medium. The isolated virus was further purified with two rounds of centrifugation at $15000\times g$ for 10 min at 4°C and then supernatants were collected and stored at -80°C . RT- PCR for (RSV) was then carried out to confirm virus identity.

2.8. Nested RT-PCR

The Nested RT-PCR was carried out for the RT-PCR positive (RSV) samples including cell culture isolate using primer pairs located on the RSV G gene (Oligomicrogen, Korea). The following primers were used [19]: G1- CCA TTC TGG CAA TGA TAA TCT C G2- GTT TTT TGT TTG GTA TTC TTT TGC GA G3- CGG CAA ACC ACA AAG TCA CAC G4- GGG TAC AAA GTT AAA CAC TTC The primers G1 and G2 were used for the first round with 20 μ L of PCR master mix (iNtRON BIOTECHNOLOGY, Korea) according to the manufacture instructions. The PCR was performed for 40 cycles, initially for 5 min at 95°C followed by, 1 min at 94°C, 1 min at 56°C and 1 min at 72°C, and finally 5 min of extension at 72°C for one cycle. For the 2nd round, 35 cycles was performed using G3 and G4 primers, initially for 5 min at 95°C followed by, 1 min at 94°C, 1 min at 56°C, 1 min at 72°C, and finally 5 min at 72°C for one cycle. The expected final Nested-PCR product was 326 bp. The Nested RT-PCR products were purified and sequenced by Macrogen's sequencing service (Amsterdam, Netherlands).

2.9. Bioinformatics Analysis

Seven consensus sequences of RSV isolates from children less than 5 years of age were successfully analyzed. The sequences were edited manually to correct possible base calling errors using BioEdit 7.0 [20], the sequences chromatogram was viewed by FinchTV1.4.0 program [21]. Then the nucleotides sequences of G genes were searched for sequences similarity using nucleotide BLAST [22]. Highly similar sequences were retrieved from NCBI and subjected to multiple sequence alignment using BioEdit software. Phylogenetic tree of G genes and their evolutionary relationship with those obtained from

database was reconstructed using the maximum likelihood online by phylogeny.fr [23]. The seven sequences have been deposited with the Gene Bank under accession numbers (AB979187, AB979188, AB979189, AB979190, AB979191, AB979192 and AB979193).

2.10. Statistical Analysis

The collected data was analyzed using statistical package for social science (SPSS) version 20, Chi square test was used, a p-value of <0.05 was considered significant.

3. Results

3.1. Detection of RSV

During the study period, 224 children patients were enrolled. Out of these, 151 children were under 2 years of age and 73 were between 2-5 years of age. RSV infections were detected in 136 (60.7%) of these patients, using DFA technique (Table 1, Figure 1).

A total of 224 swabs specimens were tested by RT-PCR, out of these RSV was detected in 44 (19.6%) patients. of which 30 (13.4%) patients were <2 years of age and 14 (6.2%) patients were 2-5 years of age (Table 1, Figure 1). There was no statistically significant association between age and RSV infection (P-value=0.903) (Table 1). Out of 44 positive samples by RT-PCR, 22 strains of RSV were isolated in Hep-2 cell line.

The figure show percentage of positive and negative RSV cases detected by direct immunofluorescence assays (DFA) and reverse transcription- polymerase chain reaction (RT-PCR).

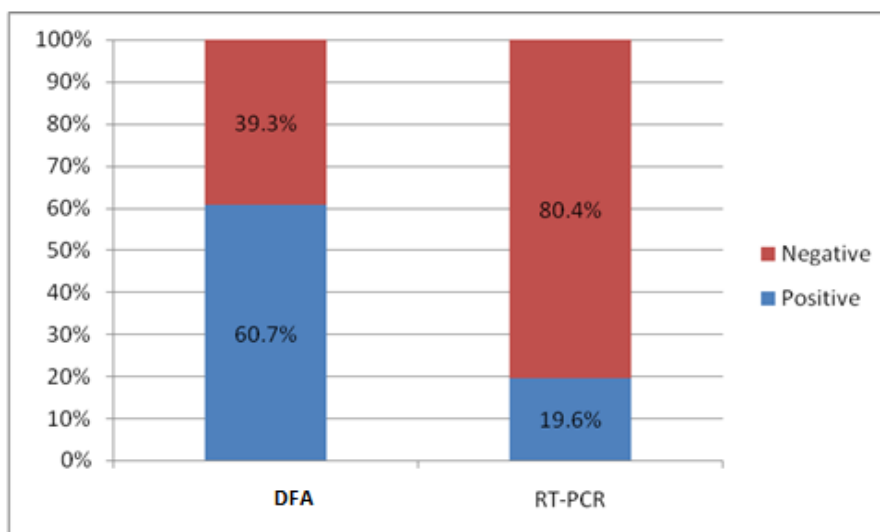


Figure 1. Detection of Respiratory Syncytial Virus infection in children by RT-PCR and DFA

Table 1. Age-related Respiratory Syncytial Virus Infection in Children in Khartoum state using DFA and RT-PCR techniques

Age group (years)	DFA		Total	RT-PCR		Total
	Positive	Negative		Positive	Negative	
<2 years	92	59	151	30	121	151
% Of Total	41.1%	26.3%	67.4%	13.4%	54.0%	67.4%
2-5 years	44	29	73	14	59	73
% Of total	19.6%	12.9%	32.6%	6.2%	26.3%	32.6%
Total	136	88	224	44	180	224
% Of total	60.7%	39.3%	100%	19.6%	80.4%	100%
P-value	0.925		-	0.903		-

3.2. Detection of RSV in 2011-2012

The prevalence of RSV in 2011 was 61 (54.5%) using DFA technique and 21 (18.7%) using RT-PCR technique.

In 2012 the RSV was detected in 75 (67%) samples using DFA and 23 (20.5%) samples using RT-PCR.

Table 2. Seasons -related Respiratory Syncytial Virus Infection in Children in Khartoum state using DFA and RT-PCR techniques

Seasons	DFA		Total	RT-PCR		Total
	Positive	Negative		Positive	Negative	
2011	61	51	112	21	91	112
%	54.5%	45.5%	100%	18.7%	81.3%	100%
2012	75	37	112	23	89	112
%	67%	33%	100%	20.5%	79.5%	100%
Total	136	88	224	44	180	224

3.3. Clinical Symptoms Associated RSV

The clinical symptoms associated with RSV in positive patients were described in Table 3 and Figure 2. RSV-positive children under 2 years of age associated with Bronchiolitis, pneumonia, Asthma and Allergy, represented 12 (27.3%), 19 (43.2%), 3 (6.8%) and 1 (2.3%), respectively. RSV-positive children in 2-5 years

of age were, 11 (25%), 2 (4.5%), 9 (20.5%), 8 (18.2%) respectively. The four clinical parameters showed significantly different rates ($p < 0.05$) in having RSV infection, (P -value = 0.017, 0.002, 0.0001, 0.0001) for Bronchiolitis, Pneumonia, Asthma and Allergy, respectively (Table 3).

Table 3. Clinical symptoms associated RSV-positive patients by RT-PCR in Khartoum state

Age group of +ve RSV children	Bronchiolitis		pneumonia		Asthma		Allergy	
	+ve samples	-ve samples	+ve samples	-ve samples	+ve samples	-ve samples	+ve samples	-ve samples
<2 years	12	18	19	11	3	27	1	29
%	27.3%	40.9%	43.2%	25%	6.8%	61.4%	2.3%	65.9%
2- 5 years	11	3	2	12	9	5	8	6
%	25%	6.8%	4.5%	27.3%	20.5%	11.4%	18.2%	13.6%
Total	23	21	21	23	12	32	9	35
%	52.3%	47.7%	47.7	52.3%	27.3%	72.7%	20.5%	79.5%
P-value	0.017		0.002		0.0001		0.0001	

P-value = (0.017, 0.002, 0.0001, 0.0001) for Bronchiolitis, Pneumonia, Asthma and Allergy, respectively, all of them showed a significant difference.

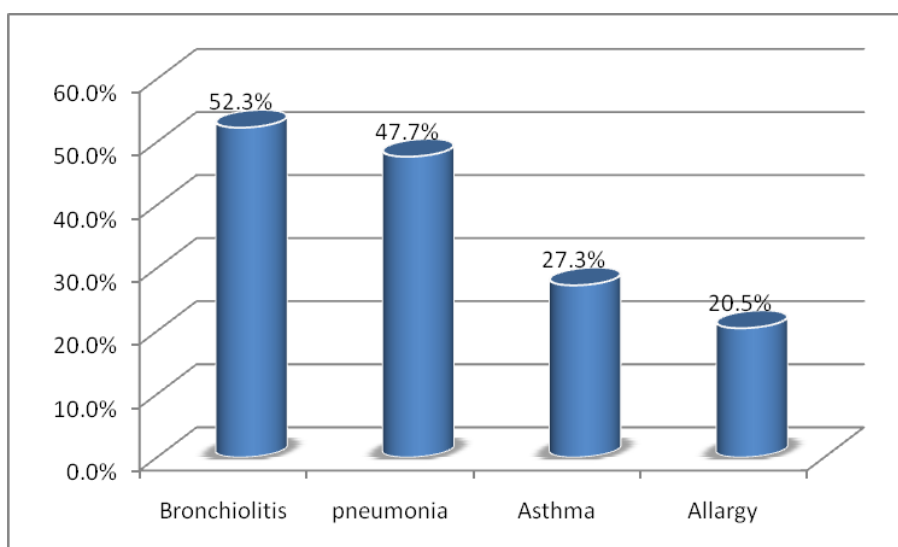


Figure 2. Distribution of positive RSV Patients according to clinical diagnosis using RT-PCR

The figure show percentage of positive RSV cases in Khartoum hospitals with clinical symptoms (Bronchiolitis, Pneumonia, Asthma and Allergy) determined by reverse transcription- polymerase chain reaction (RT-PCR).

3.4. Gender associated RSV

According to the gender, RSV was detected in 29 (12.9%) male and 15 (6.7%) female patients, using RT-PCR technique (Table 4). However our study showed that there was no statistically significant difference between males and females in having RSV infection (P -value=0.699) (Table 4).

Table 4. Respiratory syncytial virus positive cases in male and female patients by using RT-PCR

Gender	RSV		Total	P-value
	Positive	Negative		
Male	29	113	142	0.699
% Of Total	12.9%	50.4%	63.4%	
Female	15	67	82	
% Of Total	6.7%	29.9%	36.6%	
Total	44	180	224	
% Of Total	19.6%	80.4%	100%	

3.5. Nucleotide Sequence and Bioinformatics Analysis of RSV

Successful sequencing of HRSV based on G gene was done on 7 samples. Phylogenetic analysis of nucleotide sequences revealed that all analyzed RSV belonged to the RSV-A strain. The nucleotide sequences and their accession numbers in gene bank are shown in Table 5. Furthermore, on comparison with reference A2 strain (GenBank accession number M74568) and others strains from gene bank, transversionmutation was identified in

one of the Sudanese viruses (Isolate 316) at position 220 were C was replaced by A (Figure 3).

The Sudanese nucleotide sequences were compared with other sequences published in Gene Bank. Phylogenetic tree that were generated for RSV subtype-A are shown in Figure 4.

The Sudanese RSV-A strains belonged to the same clade with JX645826 A strain (Belgium) and JX131640 A strain (Saudi Arabia) (Figure 4).

Table 5. Sudanese Respiratory Syncytial virus (RSV) Sequences and Related information

Sample name	Date of collection	Age	Gender	Genebank accession number
Isolate 17	29-12-2011	4 y	M	AB979187
Isolate 127	29-01-2011	4 y	M	AB979188
Isolate 129	29-01-2011	2 y	F	AB979189
Isolate 316	16-01-2012	5 y	F	AB979190
Isolate 354	07-02-2012	2 m	F	AB979191
Isolate 372	11-02-2012	3 m	M	AB979192
Isolate 395	19-02-2012	1 y	M	AB979193

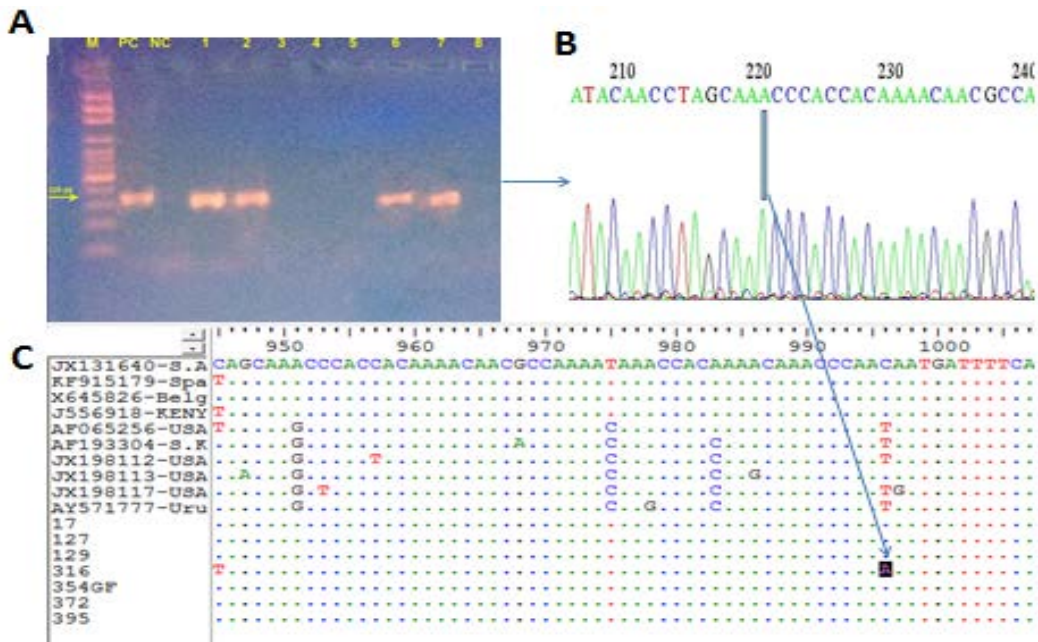


Figure 3. chromatogram and multiple sequence alignment

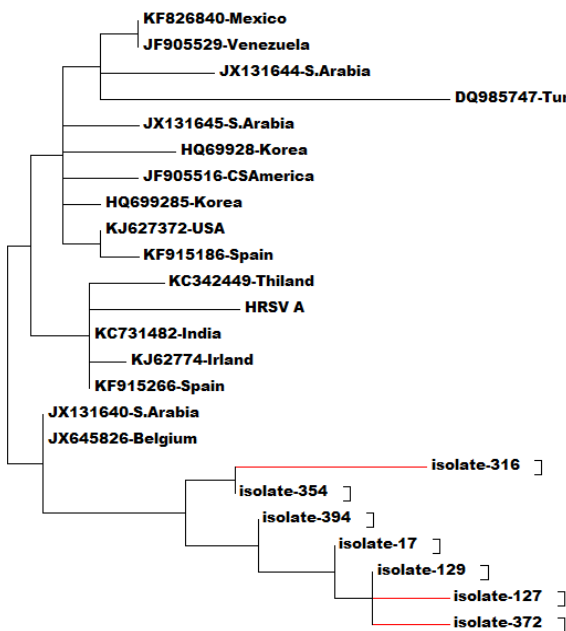


Figure 4. Phylogenetic analysis of the RSV G gene

A. PCR amplification of G gene of RSV on 1.5% agarose gel electrophoresis. Lane M DNA ladder: MW 100 bp. Lane PC; positive control. Lane NC; negative control, Lane 1, 2, 6 and 7 showing typical band size of (326 pb) corresponding to the molecular size of G genes, 3, 4, 5 and 8 negative samples. B. Sequences chromatogram of isolate 316 shown by Finch TV software. C. BioEdit multiple sequence alignment. There is transversionmutation in which C was replaced by A illustrated by the arrows.

Alignment of G gene sequences was undertaken with BioEdit software. Sequences were aligned with 17 sequences of RSV subgroup A downloaded from gene bank. The phylogenetic tree was designed using phylogeny.fr software.

4. Discussion

Over the past decade, number of studies has confirmed that RSV represents a substantial burden of acute respiratory tract illness particularly in the early years of life leading to severe morbidity and hospitalization in children [24,25,26]. Knowledge of the RSV molecular

epidemiology in Sudan is narrow. However, data collected from annual records of Ministry of Health [18] in Sudan showed that, within the ten top listed diseases causing hospitalization in infants, Pneumonia accounted for (28.3-30%) during 2004-2008. Asthma accounted for (1.8-2.5%) during 2004-2007 while acute bronchitis accounted for (2%) in 2008. The reports showed that pneumonia was the first disease leading to death during 2004, 2005 and 2006 accounting for 1126, 918, 814 deaths respectively. A recent study in Khartoum state reported that, the RSV infection was detected in 27 out of 334 patients that were negative for influenza viruses [27].

We in here, provides the first report on molecular detection and characterization of RSV in Sudan. In this study we investigated the incidence of respiratory syncytial virus (RSV) infection and characterized the RSV circulating in Khartoum state, during winter.

Two hundred and twenty four of samples were collected from children with RTI and significantly high rate of RSV infection was detected by RT-PCR, where by 44 (19.6%) were reported as positive cases. Similar result were recorded in Egypt in which, RSV was detected in 16.4% of the cases [28]. However, lower incidence were reported from Kenya where 166 out of 2143 (8%), and from India where 21 out of 200 (10%) were confirmed positive for RSV infection [29,30].

A limitation of these latter results is that there was a possibility of missing cases due to no visit to the clinic because of mild symptoms and considerable distances between villages and hospitals. The high rate of RSV in our result explains the increase in RTI cases during winter seasons in Khartoum state.

The present study showed that the prevalence rate of RSV was 136 (60.7%) using DFA and 44 (19.6%) using RT-PCR, this variation in the result between the two techniques was due to the fact that the DFA technique was less sensitive method as compared to RT-PCR that has shown improved sensitivity in the detection of RSV infection [31,32].

The current study showed that the prevalence of RSV was high in 2012 than in 2011 by the two techniques (DFA and RT-PCR), were 75 (67%) and 23 (20.5%) respectively. This result may possibly due to the significant variation in RSV outbreaks from year to year [33]. In addition, there is no awareness to prevent the RSV in Sudan.

Although the bronchiolitis is a prominent sign for clinical diagnosis of patients with RSV infection especially in infants [34]. However, our results showed that pneumonia was the most frequent clinical diagnoses in hospitalized RSV cases in age group <2 years, constituting 19 (43.2%). The Bronchiolitis was the main clinical diagnosis in RSV cases in age group 2-5 years, it were 11 (25%). This variant with the results reported by RoyaNikfar [19], in which bronchiolitis was the most frequent clinical diagnoses in children younger than 2 years old, (33%). And pneumonia was the most frequent clinical diagnoses in children from 2-5 years old, (34%). The variation in our result may be due to the fact that, acute bronchiolitis children are at risk for developing pneumonia. Statistical analysis showed significant difference in different clinical symptoms, Bronchiolitis, Pneumonia, Asthma and Allergy (P-value = (0.017, 0.002, 0.0001, 0.0001, respectively).

Various studies suggest that male children are more susceptible to severe RSV infection than females [35,36]. Parallel results were obtained in our study; in which higher percentage of RSV-infected children were males. However, statistical analysis on clinical features and hospitalization with RSV rates between male and female patients did not reveal any striking differences.

The present findings show that the RSV was of higher prevalence rate in group of <2 years 13.4%, than in 2-5 years 6.2%, These findings are in agreement with previous study in Brazil [37], in which a high rate of RSV infection was in children younger than 2 year of age. Strongly suggesting that the RSV is more likely to infect the age group <2 years [38].

Our data indicated that RSV subtype A is the predominant subtype circulating in Sudan, since all the analyzed samples were from hospitalized outpatients who came from different areas of the country. Previous studies in Taiwan and Brazil have also documented that Subtype A usually is the predominant circulating strains compared with subtype B [39,40].

Transversionmutation in G gene was identified in one of the Sudanese strains (Isolate 316). Several studies of RSV strains have described many mutations in G gene in different positions [41,42,43].

The Sudanese RSV viruses are closer to the previous described strains in Saudi Arabia, and that is mainly due to the short distance between Sudan and Saudi Arabia beside the great number of Sudanese people who travel to and from Saudi Arabia for Hajj, umrah and mostly for work. Furthermore, our viruses are also very close to Belgium viruses, a fact that is hard to explain, in view of the fact that Belgium is far than many other European countries such as France, Germany and Italy, in which other different RSV strains are circulating [44,45,46].

Moreover, no identical sequences to the Sudanese isolates could be found in Gene Bank. This is plausible because limited sequence data is available at present time on the prevalence of RSV in Africa, which are more likely to share the same genotypes.

Our earlier study on the prevalence of RSV infection in infants in Sudan showed that, RSV antibodies were more likely to be detected in re infected infants (28.8%) with respiratory infections than in first time infected infants (8.3%) (Unpublished data). Since we did not address this question in our present study, it is hard to make any comparison with our earlier study in regard to prevalence of RSV in cases of re infection.

5. Conclusion

In conclusion, this is the first report on the molecular epidemiology of RSV associated RTI in Sudan. It highlights the significance of RSV as a dominant etiologic agent contributing towards morbidity and mortality especially in young children in Sudan, and need for further work on incidence of viral pneumonias and their impact on public health. The detection and characterization of RSV infection may help to further our understanding with respect to acute respiratory infections of children.

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Conflict of Interest

All authors declare that none of them have any conflict of interest.

Authors' Contributions

SOK carried out the sample collection, virus isolation, DFA, RT-PCR, and drafted the manuscript. BS designed the alignment and phylogenetic analysis. A B M did the Statistical analysis. KAE helped in the work of virus isolation, RT-PCR and help in the drafting of the manuscript. IME and YHI supervised the work and drafting of the manuscript. All authors read and approved the final manuscript.

Ethical Clearance

The study was approved by the Ethical Review Committee (ERC) of the Ministry of Health Khartoum State, Sudan. Written consents were obtained from all parents of the patients.

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