

# Changes in Enzymes Activities of *Clarias Gariepinus* Brood Fish Exposed to Anaesthetics Metomidate

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**Abstract** The effect of anaesthetic metomidate on the enzymes activities (Alanine transaminase – ALT; Aspartate transaminase – AST; Alkaline phosphatase-ALP and Lactate dehydrogenase – LDH) in female *Clarias gariepinus* was investigated using different concentrations (0.00mL<sup>-1</sup> – control; 0.25; 0.75; 1.00; 2.00; 6.00; 8.00; 10.00 and 12.00mL<sup>-1</sup>) in triplicates. The results from the study indicated that the anaesthetic caused a concentration dependent significant increase ( $p < 0.05$ ) in the four enzymes under consideration. The highest activities in all the enzymes were observed in 12.00mL<sup>-1</sup> and the lowest in the control. The results from this work therefore suggests that the anaesthetics impair enzymes activities in the fish which was more noticeable in the fish exposed to higher concentration of 8.00-12.00mL<sup>-1</sup>. Hence caution should be exercised in the application of this anaesthetics in sedation of *C. gariepinus*.

**Keywords:** anaesthetics, aquaculture, stress, catfish, enzymes

## 1. Introduction

The intensive nature of aquaculture worldwide involves manipulation of fish. This entails some management practices such as handling, confinement, transportation and other farm management operations from the hatchery to the final commercial stage [1,2]. These procedures as crucial as they are, produce some level of disturbances [3], which can elicit a stress response leading to decreased fish performance [4] and alterations of the peripheral leucocytes distribution, haemoglobin content and red blood cell indices [5,6]. It should be noted that some very devastating effects of stress on the stock may occur during application of handling procedures in aquaculture with and without apparent warning. This then raises the question of proper monitoring of stress in fish, in order to reduce to the barest minimum the negative effects of such management procedures on fish physiology [7]. Sedation and immobilization achieved by application of anaesthetics, is seen as one of the efficient means of achieving this.

Anesthesia is generally defined as a loss of sensation through depression of the nervous system caused by an applied external agent. Anaesthetics are among important veterinary medicines employed in various hatchery operations and fish transportation. Anaesthesia is a state of unconsciousness induced in an animal by a chemical. The three levels of anaesthesia are anaesthesia (pain relief), amnesia (loss of memory) and immobilization [8]. The drugs used to achieve anesthesia usually have varying

effects in each of these areas. According to Tort et al. [9], some drugs may be used individually for these purposes or in combination with other drugs to achieve full anaesthesia. The use of anaesthetics facilitates work with fish at the research level and is required for invasive studies. These included surgical preparation for physiological investigations, where the fish must be held immobile for extended periods of time [10,11].

Anaesthetics may be local or general depending on their purpose and application, also, method and administration for each anaesthetic is fairly well defined [12], but the choice of anaesthetic depends on a number of factors. For example, if the maintenance of gill ventilation during an experimental procedure is desirable, then ketamine hydrochloride would be one possible anaesthetic [13]. In minimizing transportation stress, a light sedation brought about by low concentrations of an anaesthetic such as TMS buffered with sodium bicarbonate [14]. Anaesthetics such as MS-222, clove oil, quinaldinesulphate, and metomidate are popularly used during production procedures in fish hatcheries process [15].

Anaesthetics act with various intensity, driving fish into general anaesthesia, resulting in loss of consciousness, inhibition of reflex activity and reduced skeletal muscle movement [16]. Regardless of the agent, the process of anesthesia in fish develops in a similar way and runs in a progressive pattern, however, overdose of an anaesthetic or retaining the fish in an anaesthetic bath for too long leads to the fading of ventilation, hypoxia and finally respiration and cardiac collapse [17].

Onset of anaesthesia also caused changes in some enzyme activities of the fish [18]. A study by Trimmer et al., [19], demonstrated changes in some enzyme activities in fish, which included enzymes activity in the brain of male African catfish, *Clarias gariepinus*. This enzymes is involved in the biosynthesis of catecholestrogens that plays a role in the negative feedback mechanism of gonadal steroids on gonadotropin release. In an extensive study by Weber et al., [20], alterations were observed in some plasma enzymes, alanine transaminase (ALT), aspartate transaminase (AST), alkaline phosphatase (ALP), lactate dehydrogenase (LDH) in Indian catfish, *Clarias batrachus* exposed to various levels of an anaesthetic, 2-phenoxyethanol and metomidate in the laboratory. Velisek et al., [21] associated distortion in plasma enzyme activities of rainbow trout with the effect of anaesthetic metomidate. However, there is paucity of information in the effects of metomidate anaesthetics on the enzyme activities in *Clarias gariepinus*, hence the need to carry out this work.

## 2. Materials and Methods

A total of 66 gravid female fish ready to spawn brood fish (mean length 60.23cm  $\pm$ 0.36SD and mean weight 2.640g  $\pm$ 1.365SD) were obtained from a private fish farm, water shed, fish farms, Ru mudara, Port Harcourt, Rivers State, Nigeria and transferred in an open 100l plastic tank to the laboratory in department of Fishery and Aquatic Environment, Rivers State University of Science and Technology, Port Harcourt, Rivers State, Nigeria. They were acclimated to laboratory for a period of 10 days. After this, they were exposed to solution of anaesthetic metomidate at different concentrations (0.00 – control, 0.25, 0.50, 0.75, 1.00, 2.00, 4.00, 6.00, 8.00, 10.00, and 12.00mL<sup>-1</sup>) in 30L aquaria in triplicates for a period of 1 hour, when the fish in all exposed concentration have become immobilized [22].

Blood samples for enzyme assays and transminase activities, were collected from fish in each concentration including the control. This was done by severity the caudal pendicle, with 21 Gauge hypodermic needle and 5ml disposable syringe and transferred into heparinised bottle, this was latter centrifuge at 500rpm for 5min in (Hettich Zentrifuges, ROTINA 380) to obtain blood serum samples and stored at – 20 °C. ALT, AST, and ALP were

determined by near-infrared particle immuno-assay detection system with synchron L x 20 PRO (Beckman Counter Inc. Fullerton, CA, USA), LDH was evaluated according to Svelger et al. [23] using kits from biomerieux, France.

Data from the study was analyzed using one way analysis of variance (ANOVA) at 0.03% probability and differences among means was determined by Tuckeys multiple comparison test [24].

## 3. Results

There were no obvious signs of disease or abnormality in the physical conditions and the behaviour of the experimental fish during the experimental period. No mortality was recorded in the brood fish during the treatment. Introduction of metomidate anaesthetics in the tanks did not cause any significant change ( $P > 0.05$ ) in the water quality variables before and after the trial (Table 1). However, it caused a drop in the dissolved oxygen level and light rise in ammonia concentration.

**Table 1. Physicochemical parameters of water in the experimental tanks during trial**

Parameter	Before Trial Mean $\pm$ SD	After trial Mean $\pm$ SD
pH	6.56 $\pm$ 0.39 <sup>a</sup>	6.60 $\pm$ 0.14 <sup>a</sup>
DO (mg/L)	5.22 $\pm$ 0.78 <sup>a</sup>	4.99 $\pm$ 0.16 <sup>a</sup>
Temperature ( °C)	28.17 $\pm$ 0.71 <sup>a</sup>	28.14 $\pm$ 0.32 <sup>a</sup>
Ammonia (mg/L)	0.38 $\pm$ 0.01 <sup>a</sup>	0.40 $\pm$ 0.11 <sup>a</sup>
Nitrite (mg/L)	0.0039 $\pm$ 0.01 <sup>a</sup>	0.0043 $\pm$ 0.12 <sup>a</sup>
Sulfide (mg/L)	0.01 $\pm$ 0.01 <sup>a</sup>	0.01 $\pm$ 0.01 <sup>a</sup>

Means within the column with same superscript are not significant ( $p < 0.05$ )

The effects of anaesthetics on the plasma enzymes indicated that ALP activity in the plasma of the treated fish increased from the control 0.00mL/L (23.00  $\pm$ 2.09IU/L) and peaked (59.23  $\pm$ 1.37IU/L) at 12.00mL/L (Figure 1). Similar trends were recorded for AST and ALT (Table 2) AST activity in most of the concentrations were about twice that of ALT.

The highest values of AST, (145.66  $\pm$ 11.01U/L), ALT (76.67  $\pm$ 1.14U/L) LDH (82.66  $\pm$ 1.531U/L) were recorded fish exposed to 12.00mL/L of metomidate, while the lowest values were in all the enzymes were recoded in the control (Table 2).

**Table 2. Effects of anaesthetics (metomidate) on enzymes profiles gravid of *C. gariepinus* (mean  $\pm$ SD)**

Conc (mL/L)	Plasma Enzymes			
	ALP (IU/L)	AST (IU/L)	ALT (IU/L)	LDH (IU/L)
0.00	23.00 $\pm$ 2.09	18.54-27.46	5.75 $\pm$ 2.48 <sup>a</sup>	16.25 $\pm$ 5.98 <sup>av</sup>
0.25	31.75 $\pm$ 2.00 <sup>b</sup>	22.25 $\pm$ 1.01 <sup>c</sup>	4.50 $\pm$ 2.48 <sup>a</sup>	34.00 $\pm$ 2.91 <sup>b</sup>
0.50	36.25 $\pm$ 2.14 <sup>c</sup>	46.75 $\pm$ 8.14 <sup>d</sup>	14.25 $\pm$ 2.48 <sup>b</sup>	59.25 $\pm$ 1.91 <sup>c</sup>
0.75	38.75 $\pm$ 1.52 <sup>d</sup>	79.75 $\pm$ 4.50 <sup>db</sup>	40.25 $\pm$ 2.48 <sup>c</sup>	70.75 $\pm$ 5.13 <sup>c</sup>
1.00	42.25 $\pm$ 2.02 <sup>e</sup>	88.25 $\pm$ 2.08 <sup>b</sup>	46.50 $\pm$ 2.48 <sup>c</sup>	75.00 $\pm$ 1.00 <sup>c</sup>
2.00	44.67 $\pm$ 1.54 <sup>f</sup>	92.33 $\pm$ 2.88 <sup>e</sup>	49.67 $\pm$ 1.52 <sup>bc</sup>	75.33 $\pm$ 2.08 <sup>c</sup>
4.00	46.68 $\pm$ 1.52 <sup>f</sup>	99.99 $\pm$ 1.00 <sup>e</sup>	54.00 $\pm$ 2.01 <sup>e</sup>	76.33 $\pm$ 2.51 <sup>b</sup>
6.00	49.33 $\pm$ 1.02 <sup>db</sup>	107.33 $\pm$ 3.05 <sup>ef</sup>	60.06 $\pm$ 2.3 <sup>c</sup>	78.33 $\pm$ 1.15 <sup>b</sup>
8.00	51.66 $\pm$ 1.8 <sup>ef</sup>	115.00 $\pm$ 5.04 <sup>f</sup>	67.67 $\pm$ 1.52 <sup>f</sup>	79.01 $\pm$ 2.00 <sup>b</sup>
10.00	55.00 $\pm$ 1.00 <sup>gh</sup>	130.00 $\pm$ 5.11 <sup>g</sup>	71.33 $\pm$ 2.08 <sup>f</sup>	82.66 $\pm$ 1.53 <sup>b</sup>
12.00	59.23 $\pm$ 1.37 <sup>gh</sup>	145.66 $\pm$ 11.01 <sup>h</sup>	76.67 $\pm$ 1.14 <sup>f</sup>	82.66 $\pm$ 1.53 <sup>b</sup>

**Key:** ALP – Alkaline Phosphate, AST – Aspartate Transaminase, ALT – Alanine Transaminase, LDH – Lactate dehydrogenase  
Means within the column with same superscript are not significant ( $p > 0.05$ ).

## 4. Discussion

Transaminases are both plasma and non-plasma specific enzymes that are found in the tissues of fish and normally give information about organ dysfunction. In most teleost fish, enzyme activities affect various chemical and biological reactions in the body of the fish [25]. According to Gabriel and George [26], transamination is one principal pathway for synthesis and deamination of amino acids, enabling carbohydrate and protein metabolism during fluctuating energy demands of the organism under various adaptive conditions. Beyer *et al.*, [27] opined that alanine and aspartate transaminase activities may be utilized as sensitive markers in experimental insecticide or herbicide intoxication in teleost fish.

Alkaline phosphatase (ALP) is an enzyme present practically in all tissues especially in the cell membranes, where active transport normally takes place and has hydrolase and transphosphorylase function. ALP activity in *C. gariepinus* brood fish increased significantly ( $p < 0.05$ ) and peaked at 12.00 ml/l, similar result was recorded by Iverzen *et al.* [28], in atlantic salmon. Wagner *et al.* [29] reported an increase ( $p < 0.05$ ) in ALP activity in rainbow trout exposed to higher concentrations of clove oil (anaesthetics). Available report shows that increase or decrease in ALP activity can be used as index of hepatic parenchymal damage and hepatocytic necrosis [30]. Therefore an increase in the ALP enzyme activities reflects activities in inflammation and necrosis of hepatic cells.

The activities of ALT and AST increased correspondingly, as the concentrations of the metomidate increased, a pattern noted by Velisek and Svobodova, [31], in common carp after exposure to higher concentrations of MS-222. Also, Iwama and Aerkerman, [32], observed the same result in rainbow trout exposed to increasing concentration of MS222 and clove oil. The disorder in plasma enzymes could be from of hepatic injury as a result of reactive metabolites from xenobiotic metabolism in the liver [33]. A concentration dependent activities of LDH noted in fish has previously been reported by Spotte *et al.*, [34] in sea water adapted mummichog exposed to 2-phenoxyethanol in the laboratory, also, Ferreira *et al.* [35], in rainbow trout exposed to benzocaine hydrochloride and Guo *et al.* [36], in *Xiphophorus maculatus* treated with MS 222 in the hatchery. The trend suggests an increase in the glycolytic process due to the lower metabolic rate as a result of the effect of anaesthetics. This is because the level of tissue lactate content acts as an indicator of anaerobic respiration as this enhances the animal tolerance to internal hypoxic condition [37].

## 5. Conclusion

The activities in the plasma enzymes of *C. gariepinus* was impaired by anaesthetics metomidate, which was more pronounced at higher concentration of (8.00 – 12.00 ml/L<sup>-1</sup>). Therefore, caution should be exercise in the application of this anaesthetics in sedation of *C. gariepinus*.

## References

- [1] Angelids, P; Baudin – Laurecin, T; and Youinov, S. (1987). Stress in rainbow trout salmon *geivrdineri*, effects upon phagocytes and susceptibility to *Aeromonas salmonicida* *J. Fish. Biol.* 31:113-122.
- [2] Gabriel, U. U.; P. E. Anyanwu and Akinrotimi O. A. (2007a). Blood characteristics associate with continent stress in Black chin Tilapia, *Sarotherodon melanotheron* *J. Fish. Intl.* 2(2):186-189.
- [3] Pickering, A. D. (1981). *Stress and Fish*. Academic Press New York. 367pp.
- [4] Cubero, L. and Molinero, A. (1997). Handling confinement and anesthetic exposure induced changes in the blood and tissue immune characteristics of gilthead sea bream. *Dis. Aquat. Org.* 31:89-94.
- [5] Ainsworth, A. J; C. Dexlag and P. R. Waterstrat (1991). Changes in peripheral blood leukocyte percentages and function of new cells in stressed channel catfish. *J. Aquat Anim. Health* 3:41-47.
- [6] Gabriel, U. U; P. E. Anyanwu and A. O. Akinrotimi (2007b). Haematological profile of black-chinned tilapia *Sarotherodon melanothem* from Buguma Creek, Niger Delta. *Agric. J.* 2(3): 384-387.
- [7] Akinrotimi, O. A., Gabriel, U. U., Anyanwu, P. E. and Anyanwu, A. O. (2007). Influence of sex, acclimation methods and period on haematology of *Sarotherodon melanotheron*. *Res. J. Biol. Sci.* 2(3): 348-352.
- [8] Otruno J; Esteban U. A. and Meseguer, J. (2002). Lack of effect of combining different stressors on innate immune response of sea bream (*Spanus aurata* L). *Vet. Immunol Immunopathol* (84):17-27.
- [9] Tort, L; Puigcerver, M., Crespo, S. and Padros F. (2002). Cortisol and haematological response in sea bream and trout subjected to the anaesthetics clove oil and 2-phenoethanol. *Aquaculture Research* 33:907-910.
- [10] Akinrotimi, O.A. Opara, J.Y and Ibemere, I.F. (2012). Changes in haematological parameters of Tilapia *guineensis* exposed to different water pH environment *Innovations in Science and Engineering* 2:9-14
- [11] Velisek J. and Svobodova Z. (2004). Anesthesia of rainbow trout (*Oncorhynchus mykiss*) with 2-phenoxyethanol: acute toxicity and effects on biochemical blood profile. *Acta Vet. Brno.* 73:378-384.
- [12] Velisek, J. and Svobodova, Z., Piackova V., Groch L., Nepekchalovah (2003). Effects of clove oil anaesthesia in common. *Carp. Vet. Med.* 50:269-274.
- [13] Graham, M. S. and Iwama, G. K. (1990). The physiologic effects of the anaesthetic, ketamine hydrochloride on two salmonid species. *Aquaculture* 90:323-332.
- [14] Bell, G. R. (1987). Distributions of transaminases (aminotransferases) in the tissue of Pacific salmon (*Oncorhynchus mykiss*) with emphasis on the properties and diagnostic use of glutamic-oxalacetic transaminase *J. Fisheries. Res. Board Can.* 25:1247-1268.
- [15] Holloway A. C., Keene J, Noakes, D. G; Moccia, R. D. (2004). Effects of clove oil and Ms-222 on blood hormones profiles in rainbow trout *Oncorhynchus mykiss*. *Aquacult. Res.* 35:1025-1030.
- [16] Hamoakova, J; Lepicova, A; Kozakp; Stupla, Z. Kouril, J and Lepic P. (2004). The efficacy of various anaesthetics in tench (*Tinca tinca*) related to water temperature *Vet. Med.* 49:467-472.
- [17] Gomes, L. C.; Duran E; Gasquez; S. Mosotz and Roncero, V. (2003). Effect of fish density during transportation on stress and mortality of juvenile tambaqui *Colossoma macropomum*. *J. World Aquacult. Soc.* 34(1):76-84.
- [18] Wagner, E., Arudt. R. and Hilton B. (2002). Physiological stress responses, egg survival and sperm mobility for rainbow trout brood stock anaesthetized with clove oil tricaine methane sulfonate or carbon dioxide. *Aquaculture* 211:353-366.
- [19] Trimmers, R. J. Granneman, J. C. Lambert, J. G. and Vanoordt, P. G. (1988). Estrogen-2-hydroxylase in the brain of the male Africa catfish, *Clarias gariepinus*. *Gerontology and Comparative Endocrinology* 72(2):190-203.
- [20] Weber, R.A., Peleteriro, J.B., Garcia, Martin I.O. and Aldegonde, M. (2009). The efficacy of 2-phenoxyethanol metomidate, clove oil and MS-222 as anaesthetics agents in the *Senegalese Sule*. *Aquaculture* 288:147-150.
- [21] Velisek, J. and Svobodova, Z. and Piakova V. (2005). Effects of clove oil anesthesia on rainbow trout (*Oncorhynchus mykiss*). *Acta Vet Brno.* 74:139-146.

- [22] Velisek, J., Wlasow T., Gomulka, P. Z., Svobodova and L. Novotry (2007). Effects of 2-phenoxyethanol anaesthesia on sheatfish *Silvanus glanis*. *Vet Med.* 52(3):103-110.
- [23] Sveliger, Y.; E.O Orve and N. Uner (2004). Ebaluation of Etioazole toxicity in the liver of *Oreochromis Niloticus* Pest. *Biochem Physiol* 78:1-8.
- [24] Wahua, T. A. T. (1999). *Applied Statistics for scientific studies*. Afrika link books. Aba,+ Nigeria. 365pp.
- [25] Oruc, E. O. and Uner, N. (2001). Marker enzyme assessment in the liver of *Cyprivus capio*. *J. Biochem. Mol. Toxicol* 16:182-188.
- [26] Gabriel, U. U and George, A. D. I. (2005). Plasma enzymes in *clarias gariepinus* exposed to chronic levels of round-up (Glycophosphate). *Env. Ecol.* 23:271-276.
- [27] Iverzen, M; Finstand, B; Mckinley, R. S; Eliassen, R. A. (2003). The efficacy of metomidate, clove oil, Agri-S and benzoak as anaesthetics in Atlantic Salmon stress reducing capacity. *Aquaculture* 221:549-566.
- [28] Beyer, J. Sanduik M; Hylland, K; Fjeld, E; Egaas, E; Aas E; Skare, J. U; Goksoy, A. (1996). Contaminant accumulation and biomarker responses in flounder (*Plantichthys flesus*) and Atlantic cod (*Cadus morhua*), exposed by caging to polluted sediments in Sotjorden, Norway. *Aquat. Toxicol.* 36:75-98.
- [29] Kildea, M.A. Allan, G.L. and Robert, E.K (2004). Accumulation and clearance of the anaesthetics clove oil and Aquis from the edible tissue of silver perch. *Aquaculture* 232:265-277.
- [30] Begum, G. (2004). Carbofuran Insecticide induced biochemical alterations in liver and muscle tissues of fish, *Clarias batrachus* and recovery response. *Aquatic Toxicology* 66(1):83-92.
- [31] Velisek J., and Svobodova. Z. (2004b): Anesthesia of rainbow trout, *Oncorhynchus mykiss*. *Acta Veterinaria Brno*, (74): 139-146.
- [32] Iwama G. K. and Ackermen, P. A. (1994). *Anaesthesia*. Pp.1-15 In: *Biochemistry and molecular biology of fishes*. Vol 3(eds P.W. Hochachka and T. P. Mommsen) Amsterdam; Elsevier Science B. V.
- [33] Levavi-sivan, B. and Avita, A. (2005). Sequence analysis, endocrine regulation and signal transduction of reactive cholesterol pools in mitochondria isolated from gonads of male goldfish (*Carassius auratus*). *Gen. Comp. Endocrinol.* 142:67-73.
- [34] Spotte, S. Bubucis, P. M. and Anderson, G. (1991). Plasma cortisol response of seawater-adapted mummichogs (*Fundulus heteroclitus*) during deep MS-222 anesthesia *Zool. Biol.* 10:75-80.
- [35] Ferreira, J. T; Shoonbee, H. J. and Smith G. L. (1984). The uptake of the anesthetic benzocaine hydrochloride by the gills and skin of three freshwater fish species. *Journal of fish Biology* (25)35-41.
- [36] Guo F. C. Teo L. H and Chea T. W. (1992). *Effects of anaesthetics on plasma cortisol and lactic acid levels in Platys (Xiphophorus maculatus)*. Bulletin of Faculty of Science Natural University, Singapore (12):30-33.
- [37] Celik, E. S. (2004). Blood chemistry (electrolytes lipoproteins and enzymes) values black scorpion fish (*Scorpaenia porcus*) in the Dardanelles. *Turkey J. Biological Sci.* 4(6):716-719.